# Discrimination of Two Tetragonula (Apidae: Meliponini) Species in Thailand using Geometric Morphometric Analysis of Wing Venation 

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#### Abstract

Geometric morphometrics is a novel powerful tool for the identification of bee species and subspecies. An application of this method is presented to discriminate two stingless bee species, Tetragonula pagdeni Schwarz, 1939 and Tetragonula fuscobalteata Cameron, 1908, in Thailand based on forewing venation. Nine landmarks were measured from the digitized right forewing of 200 workers (10 bees per colony). Eleven colonies of T. pagdeni and nine colonies of T. fuscobalteata collected from several locations in Thailand were determined. The positions of the groups in the principal component analysis plots revealed that the species are clearly distinct. Multivariate analysis of variance ( $\alpha=0.01$ ) showed a significant difference between the bees of the sibling species, producing a high degree of accuracy $(96.50 \%)$ in the identification of any individual bee of either species. Therefore, it is proposed that geometric morphometric analysis of single wings might be an appropriate first step in the analysis and identification of stingless bees and other insects.


Keywords: stingless bees, geometric morphometrics, wing venation pattern, Tetragonula pagdeni, Tetragonula fuscobalteata

## INTRODUCTION

The importance of stingless bees as effective pollinators of many plant species in natural and agro-ecosystems has been well documented (Heard, 1999; Amano et al., 2000; Slaa et al., 2000; Biesmeijer et al., 2006; Klein et al., 2007; Quezada-Euán et al., 2012), and they seem to be good candidates for applications as commercial pollinators (Slaa et al., 2006; Theeraapisakkun et al., 2010). However, there is increasing evidence
of declines in stingless bee populations (Potts et al., 2010; Francoy et al., 2011), mainly affected by habitat deterioration (Biesmeijer et al., 2006; Winfree et al., 2009; Williams et al., 2010). Thus, the accurate identification of stingless bee species is urgently needed to investigate their biodiversity (Francoy et al., 2009; May-Itzá et al., 2012).

Stingless bees (Meliponini) are one of the most morphologically and behaviorally abundant and diverse species among the corbiculate bees (Apini, Bombini and Meliponini) (Michener,

[^0]2007; Rasmussen and Cameron, 2010). Unlike the Apini, with only 11 species in the single genus Apis (Lo et al., 2010), stingless bees are a large and diverse taxon of more than 400 species comprising some 60 genera (Michener, 2007; Moure et al., 2007), many of which are poorly known (Rasmussen and Cameron, 2010). In some cases, these diverse species suggest the possible existence of many species complexes in the Meliponini (Michener, 2007; May-Itzá et al., 2012). Thus, the identification of stingless bee species remains difficult and requires analysis by an experienced taxonomist (Dollin et al., 1997; Theeraapisakkun et al., 2010).

Various molecular methods, including analyses of mitochondrial and nuclear DNA sequence polymorphism, have been widely used to discriminate among bee species and subspecies (Francisco et al., 2001; Whitfield et al., 2006; Ramirez et al., 2010; Rasmussen and Cameron, 2010; Theeraapisakkun et al., 2010; QuezadaEuán et al., 2012). However, these methods require relatively expensive reagents, specialized personal knowledge and a well-equipped laboratory (Francoy et al., 2008).

Traditional morphometric analysis, which is based on multiple measurements of various body parts across many individuals, has recently become widely used for identifying bee species and subspecies (Daly et al., 1982; Chaiyawong et al., 2004; Radloff et al., 2005; Francoy et al., 2006; Rattanawannee et al., 2007; Andere et al., 2008; Özkan et al., 2009), because of its high practicability and low cost (Francoy et al., 2008). However, traditional morphometric analysis requires time-consuming specimen preparation and measurement procedures (Francoy et al., 2006). Moreover, the complex shape of an organism cannot easily be summarized by using linear measurements as in traditional morphometrics (Pavlinov, 2001; Aytekin et al., 2007).

Instead of distance and angle measurements, geometric morphometrics uses
coordinates of homologous landmarks, which offers superior discrimination capabilities to traditional morphometry (Bookstein, 1997; Azidah et al., 2000; Villemant et al., 2007). Importantly, geometric morphometric analysis can exclude the effects of size as it uses the relative positions of the plotted landmarks. Therefore, environmental influences, for instance nutrition and parasitism, can be reduced or removed (Villemant et al., 2007; Francoy et al., 2011).

The geometric morphometric analysis of wings has been applied for the identification of bee specimens to genera, species, subspecies and even populations (Mendes et al., 2007; Francoy et al., 2008, 2011; Kandemir et al., 2009). For example, Francoy et al. (2006) showed that a single wing cell carries enough information for the identification of the three racial groups of Apis mellifera (Africanized, Italian and Carniolan) with a fidelity level of nearly $99 \%$ of the individuals. Francisco et al. (2008) showed that a geometric morphometric analysis of wing venation can be used to resolve the taxonomic identification of cryptic species in Plebeia remota. Francoy et al. (2009) reported that landmarks based on the geometric morphometric analysis of the wing shape of males and workers were sufficient to discriminate among five stingless bee species and that the sex of individuals does not impede identification. Moreover, Villemant et al. (2007) also reported that applying geometric morphometrics to wing venation could resolve a complex case of four sibling parasitoid wasp species in the genus Eubazus (Hymenoptera, Braconidae).

Even though geometric morphometric analysis is a simple methodology for which only a stereomicroscope, a digital camera and a computer are needed (Francoy et al., 2011), it gives clear results to answer the questions that are normally resolved using molecular methods (Francoy et al., 2008, 2011).

Stingless bees in the genus Tetragonula comprise the largest and most widespread group in the Indopacific area, including some of the
commonest species (Sakagami, 1978). Some stingless bee species are easily distinguished from others. But some species require critical studies, especially those that appear to be cryptic species as is the case for Tetragonula pagdeni and Tetragonula fuscobalteata. They are very difficult to distinguish because of the great overlap in size so that traditional morphometrics are inadequate to identify them. Therefore, the objective of this study was to apply a geomorphometric method using the characteristics of the forewings of workers to identify two sibling stingless bee species in Thailand, T. pagdeni Schwarz, 1939 and T. fuscobalteata Cameron, 1908 (Sakagami and Khoo, 1987; Theeraapisakkun et al., 2010). The identification accuracy is discussed and whether or not employing such geometric morphological
techniques will be an alternative method with greater accuracy and relevance to distinguish between these two taxonomically problematic species.

## MATERIAL AND METHODS

## Sample collection sample preparation and measurement

Adult workers of 11 colonies from 8 locations of T. pagdeni and 9 colonies from 6 locations of T. fuscobalteata were sampled (Table 1, Figure 1). At least 30 workers from each colony of the two species were collected directly at the nest entrance of each colony and stored in $75 \%$ ethanol. Ten workers from each colony were randomly selected for dissection. The

Table 1 Collection of two stingless bee species from Thailand.

| No. | Sampling location | Coordinates | T. pagdeni | T. fuscobalteata |
| :---: | :---: | :---: | :---: | :---: |
| 1 | Chiang Mai | $18^{\circ} 53^{\prime} 17.16^{\prime \prime} \mathrm{N}$ | 1 colony | 1 colony |
|  |  | $98^{\circ} 51^{\prime} 41.96^{\prime \prime} \mathrm{E}$ |  |  |
| 2 | Maha Sarakham | $16^{\circ} 12^{\prime} 09.42^{\prime \prime} \mathrm{N}$ | 1 colony | - |
|  |  | $103^{\circ} 16^{\prime} 59.27^{\prime \prime} \mathrm{E}$ |  |  |
| 3 | Nakhon Sawan | $16^{\circ} 00^{\prime} 36.88^{\prime \prime} \mathrm{N}$ | - | 1 colony |
|  |  | $99^{\circ} 54^{\prime} 58.70^{\prime \prime} \mathrm{E}$ |  |  |
| 4 | Nakhon Sawan | $15^{\circ} 58^{\prime} 37.01^{\prime \prime} \mathrm{N}$ | - | 1 colony |
|  |  | $99^{\circ} 54^{\prime} 17.23^{\prime \prime} \mathrm{E}$ |  |  |
| 5 | Bangkok | $13^{\circ} 44^{\prime} 13.79^{\prime \prime} \mathrm{N}$ | 1 colony | - |
|  |  | $100^{\circ} 31^{\prime} 48.40^{\prime \prime} \mathrm{E}$ |  |  |
| 6 | Chanthaburi | $12^{\circ} 30^{\prime} 43.99^{\prime \prime} \mathrm{N}$ | 1 colony | 2 colonies |
|  |  | $102^{\circ} 10^{\prime} 11.40^{\prime \prime} \mathrm{E}$ |  |  |
| 7 | Ratchaburi | $13^{\circ} 35^{\prime} 24.73^{\prime \prime} \mathrm{N}$ | 3 colonies | 3 colonies |
|  |  | $99^{\circ} 30^{\prime} 27.12^{\prime \prime} \mathrm{E}$ |  |  |
| 8 | Samut Songkhram | $13^{\circ} 23^{\prime} 13.49^{\prime \prime} \mathrm{N}$ | 2 colonies | - |
|  |  | $99^{\circ} 57^{\prime} 11.59^{\prime \prime} \mathrm{E}$ |  |  |
| 9 | Chumphon | $10^{\circ} 29^{\prime} 46.75^{\prime \prime} \mathrm{N}$ | - | 1 colony |
|  |  | $99^{\circ} 08^{\prime} 47.03^{\prime \prime} \mathrm{E}$ |  |  |
| 10 | Chumphon | $10^{\circ} 30^{\prime} 00.11^{\prime \prime} \mathrm{N}$ | 1 colony | - |
|  |  | $99^{\circ} 09^{\prime} 42.59^{\prime \prime} \mathrm{E}$ |  |  |
| 11 | Chumphon | $10^{\circ} 28^{\prime} 46.85^{\prime \prime} \mathrm{N}$ | 1 colony | - |
|  |  | $99^{\circ} 12^{\prime} 01.68^{\prime \prime} \mathrm{E}$ |  |  |
| Total |  |  | 11 colonies | 9 colonies |

right forewing of each bee sample was removed, slide-mounted and photographed with a digital camera on a stereomicroscope. From a total of 200 wings, 110 wings of $T$. pagdeni workers and 90 wings of T. fuscobalteata workers were measured. Nine homologous landmarks, which are the most easy observable and often adopted for insect identification, on the intersections of forewing veins were manually plotted (Figure 2) using the tpsDig2 version 2.16 software (Rolhf, 2005a). The Cartesian coordinates of the landmarks were then Procrustes aligned and a partial warps analysis was performed using the tpsRelw version 1.49 software (Rolhf, 2005b). The mean configuration of the bees from a colony was used as a comparative parameter and the analysis was made at the colony level (Francisco et al., 2008; Francoy et al., 2011).

## Data analysis

A principal component analysis (PCA) was performed using the Cartesian coordinates of each homologous landmark after alignment (Amssalu et al., 2004; Francoy et al., 2006, 2009, 2011) ). A stepwise analysis was carried out to determine classification functions, followed by a canonical analysis. Then a cross-validation test was performed on $10 \%$ of randomly selected individuals as unknowns (Francisco et al., 2008; Rattanawannee et al., 2010, 2012), and the data used to calculate and check the accuracy of the equations in identifying the colonies (Francoy et al., 2011). The square Mahalonobis distances were also calculated between the centroids of the bee sample groups (Francoy et al., 2009; Francoy et al., 2011). After extraction of the measures, all statistical analyses were performed using SPSS


Figure 1 Two stingless bee species collection sites in Thailand. Numbers correspond to those in Table 1

Version 10.0 (1999; SPSS; Chicago, IL, USA).

## RESULTS

The nine anatomical landmarks (Figure 2) generated 14 relative warp measurements for each of the wing specimens. The four highest relative warps explained $71.96 \%$ of the group's variability, with $38.03 \%, 12.63 \%, 11.58 \%$ and $9.72 \%$ being explained by the first, second, third and fourth relative warps, respectively.

Five principal factors with eigenvalues greater than one were extracted from the PCA analyses. Together, the five components were able to explain $68.07 \%$ of the data set variability. The first two main principal components explained $41.06 \%$ of the variability. The coordinate variable 7 x mostly influenced the first factor and explained $25.22 \%$ of the variability among the groups, whereas the coordinate variable 1y mostly influenced the second factor and explained $15.85 \%$ of the variation.

The positions of the groups in the PCA plots of factor $1(25.22 \%)$ and factor 2 ( $15.85 \%$ ), show that the species groups are well distinguished (Figure 3), with the T. pagdeni samples placed mainly in the upper left-hand quadrant and T. fuscobalteata samples mainly in the lower right-hand quadrant of the plot so that these two
sibling species of Tetragonula species are clearly separated (Figure 3).

Multivariate analysis of variance of the measures demonstrated that the stingless bee groups were significantly different (Wilk's $\lambda=$ $0.209 ; P<.001$ ). The linear discriminant analysis of the data showed that the centroid sizes of the groups were significantly different $(P<.001)$ from each other.

From the 200 wings, seven were misclassified in the respective group (Figure 4), five from T. pagdeni and two from T. fuscobalteata. The cross-validation test using $10 \%$ of the individuals as unknowns correctly identified $96.5 \%$ of the individuals to each respective group.

## DISCUSSION

In view of the fact that the two species, T. pagdeni and T. fuscobalteata have always been difficult to identify, to achieve this end was a first priority. Inasmuch as the bees can now be clearly separated, it is only as of now possible to pursue information on the basic biological differences between them and, ultimately, to develop suitable management techniques for their future use in Thailand.


Figure 2 Right forewing of a T. pagdeni worker. The white circles indicate the respective position of each of the plotted landmarks.

In recent years, the geometric morphometric analysis of wings has been used to discriminate hymenopteran species, subspecies and populations including wasps (Baylac et al., 2003; Villemant et al., 2007; Abbasi, 2009), bumble bees (Francoy et al., 2006, 2008; Aytekin et al., 2007), honey bees (Andere et al., 2008;

Tofilski, 2008; Rattanawannee et al., 2010, 2012;) and stingless bees (Mendes et al., 2007; Francisco et al., 2008; Francoy et al., 2009, 2011). Tofilski et al. (2008) showed that the geometric analysis of forewing venation is a more powerful tool than standard morphometric analysis (using the distance and angle of various parts of the


Figure 3 Scatterplot of the two most influential factors (variable 7x and 1y, respectively) from principal component analysis of workers of two stingless bee species (Tetragonula pagdeni and $T$. fuscobalteata) measured for nine landmarks.


Figure 4 Canonical distributions of the individuals from two stingless bee species, Tetragonula pagdeni and T. fuscobalteata, based upon measurements extracted from forewing venation pattern.
body) for the discrimination of subspecies of A. mellifera. Furthermore, Francoy et al. (2011) demonstrated that the geometric morphometry of the wing venation is sensitive enough to be used for discrimination between Mexican and Central American populations of Melipona beecheii. This result was supported by a study of internal transcribed spacer (ITS) ribosomal DNA sequence-based analysis, which revealed that $M$. beecheii from Mexico presents an ITS1 haplotype (ITS1-A) different from the bees from Central America (ITS1-B and ITS1-C) (May-Itzá et al., 2012).

Sibling species are expected to reveal high morphological similarity (Aytekin et al., 2007); however, some morphological differences that allow them to be distinguished can be found when morphometric analysis is used (Moraes et al., 2004; Aytekin et al., 2007;).

Among 32 stingless bee species that have been recorded in Thailand (Schwarz, 1939; Sakagami et al., 1985; Michener and Boongird, 2004; Klakasikorn et al., 2005), some sibling species such as T. pagdini, T. laeviceps, and T. fuscobulteata are sympatric species which often complicates any preliminary identification based on the geographic distribution and basic morphological characteristics (Sakagami and Khoo, 1987; Starr and Sakagami, 1987). Even though the species diagnostic CUTP1 SCAR marker for identification has been successfully developed for T. pagdeni (Thummajitsakul et al., 2010), the limit of technical skill and cost in species identification between species still remain. The geometric morphometric analysis performed in the current study on T. pagdeni and T. fuscobalteata demonstrated that this method is efficient for correctly assigning individual colonies to their species. The results demonstrated that the forewing shapes show significant differences between these two stingless bee species, as has also been found for bumblebees (Aytekin et al., 2007), wasps (Villemant et al., 2007), Diptera (Moraes et al., 2004) and Hemiptera (Gumiel et al., 2003).

The current results are in close agreement with those of Francoy et al. (2009) who showed that wing venation patterns alone are sufficient to classify five sibling species of stingless bees. They made geometric morphometric analyses of the forewings of male and workers of Nannotrigona testaceiornis, Melipona quadrifasciata, Frieseomelitta varia, Scaptotrigona depilis, and Plebeia remota and showed that the patterns of males and workers from the same species were more similar than the patterns of individuals of the same sex from different species. The origins of the wing shape differences are unclear at present (Aytekin et al., 2007; Francoy et al., 2009). In the case of stingless bees, there can be some adaptive constraints such as foraging behavior, mating behavior, flying mechanisms and pollen loads (Aytekin et al., 2007).

The current study was able to discriminate between two stingless bee species based on wing information using a digitized wing image, which greatly facilitated and accelerated analysis. It was concluded that measurements of a small part of the entire bee body are sufficient to discriminate among stingless bee species. This methodology is simple and could be extended to finer identifications among species of bees with the addition of future landmarks. Moreover, computer programs to assist in the morphometric analysis of the wing might be a useful implementation in biodiversity and conservation studies.

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## LITERATURE CITED

Abbasi, R.M., M. Abbasi and B. Kiabi. 2009. Gemetric morphometric study of populations of the social wasp, Polistes dominus (Christ, 1791) from Zanjan province, north-west Iran. N. Z. J. Zool. 36: 41-46.

Amano, K., T. Nemato and T.A. Heard. 2000. What are stingless bees, and why and how to use them as crop pollinators? A review. Jpn. Agric. Res. Q. 34: 183-190.
Amssalu, B., A. Nuru, S.E. Radloff and H.R. Hepburn. 2004. Multivariate morphometric analysis of honeybees (Apis mellifera) in the Ethiopian region. Apidologie 35: 71-81.
Andere, C., C. Garcia, C. Marinelli, R. Cepeda, E.M. Rodriguez and A. Palacio. 2008. Morphometric variables of honeybees Apis mellifera used in ecotypes characterization in Argentina. Ecol. Model. 214: 53-58.
Aytekin, A.M., M. Terzo, P. Rasmont and N. Cagatay. 2007. Landmark based geometric morphometric analysis of wing shape in Sibiricobombus Vogt (Hymenoptera: Apidae: Bombus Latreille). Ann. Soc. Entomol. Fr. (N.S.) 43: 95-102.

Azidah, A.A., M.G. Fitton and D.L.J. Quicke. 2000. Identification of the Diadegma species (Hymenoptera: Ichneumonidae, Campopleginae) attacking the diamondback moth, Plutella xylostella (Lepidoptera: Plutellidae). Bull. Entomol. Res. 90: 375389.

Baylac, M., C. Villemant and G. Simbolotti. 2003. Combining geometric morphometrics with pattern recognition for the investigation of species complexes. Biol. J. Linnean Soc. 80: 89-98.
Biesmeijer, J.C., S.P.M. Roberts, M. Reemer,
R. Ohlemuller, M. Edwards, T. Peeters, A.P. Schaffers, S.G. Potts, R. Kleukers, C.D. Thomas, J. Settele and W.E. Kunin. 2006. Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands. Science 313: 351-354.
Bookstein, F.L. 1997. Morphometric Tools for Landmark Data: Geometry and Biology. Cambridge University Press. Cambridge, UK. 435 pp .
Chaiyawong, T., S. Deowanish, S. Wongsiri, H.A. Sylvester, T.E. Rinderer and L.D. Guzman. 2004. Multivariate morphometric study of Apis florea in Thailand. J. Apic. Res. 43: 123-127.
Daly, H.V., K. Hoelmer, P. Norman and T. Allen. 1982. Computer assisted measurement and identification of honey bees. Ann. Entomol. Soc. Am. 75: 591-594.
Dollin, A.E., L.J. Dollin and S.F. Sakagami. 1997. Australian stingless bees of the genus Trigona (Hymenoptera: Apidae). Invert. Taxo. 11: 861-896.
Francisco, F.O., D. Silvestre and M.C. Arias. 2001. Mitochondrial DNA characterization of five species of Plebeia (Apidae: Meliponini): RFLP and restriction maps. Apidologie 32: 323-332.
Francisco, F.O., P. Nunes, T.M. Francoy, D. Wittmann, V.L. Imperatriz-Fonseca, M.C. Arias and E.D. Morgan. 2008. Morphometrical, biochemical and molecular tools for assessing biodiversity: An example in Plebeia remota (Holmberg, 1903) (Apidae, Meliponini). Insectes Soc. 55: 231-237.
Francoy, T.M., M.L. Grassi, V.L. ImperatrizFonseca, W.D.J. May-Itzá and G. QuezadaEuán. 2011. Geometric morphometrics of the wing as a tool for assigning genetic lineages and geographic origin to Melipona beecheii (Hymenoptera: Meliponini). Apidologie 42: 499-507.
Francoy, T.M., R.A.O. Silva, P. Nunes-Silva, C. Menezes and V.L. Imperatriz-Fonseca.
2009. Gender identification of five genera of stingless bees (Apidae, Meliponini) based on wing morphology. Genet. Mol. Res. 8: 207-214.
Francoy, T.M., D. Wittmann, M. Drauschke, S. Muller, V. Steinhage, M.A.F. BezerraLaure, D. De Jong and L.S. Goncalves. 2008. Identification of Africanized honey bees through wing morphometrics: Two fast and efficient procedures. Apidologie 39: 488-494.

Francoy, T.M., P.P.R. Prado, L.S. Goncalves, L.D. Costa and D. De Jong. 2006. Morphometric differences in a single wing cell can discriminate Apis mellifera racial types. Apidologie 37: 91-97.
Gumiel, M., S. Catala, F. Noireau, A. Rojas de Arias, A. Garcia and J.P. Dujardin. 2003. Wing geometry in Triatoma infestans (Klug) and T. melanosoma Martinez, Olmedo and Carcavallo (Hemiptera: Reduviidae). Syst. Entomol. 28: 173-179.
Heard, T.A. 1999. The role of stingless bees in crop pollination. Ann. Rev. Entomol. 44: 183-206.
Kandemir, I., M.G. Moradi, B. Ozden and A. Ozkan. 2009. Wing geometry as a tool for studying the population structure of dwarf honey bees (Apis florea Fabricius 1876) in Iran. J. Apic. Res. 48: 238-246.
Klakasikorn, A., S. Wongsiri, S. Deowanish and O. Duangphakdee 2005. New record of stingless bees (Meliponini: Trigona) in Thailand. Nat. Hist. J. Chulalongkorn Univ. 5: 1-7.
Klein, A.M., B.E. Vaissière, J.H. Cane, I. SteffanDewenter, S.A. Cunningham, C. Kremen and T. Tscharntke. 2007. Importance of pollinators in changing landscapes for world crops. Proc. R. Soc. London B. Bio. Sci. 274: 303-313.

Lo, N., R.S. Gloag, D.L. Anderson and B.P. Oldroyd. 2010. A molecular phylogeny of the genus Apis suggests that the Giant Honey Bee of the Philippines, $A$. breviligula Maa, and the Plains Honey Bee of southern India, A. indica

Fabricius, are valid species. Syst. Entomol. 35: 226-233.
May-Itzá, W.de J., J.J.G. Quezada-Euán, R. Ayala and P. de la Rua. 2012. Morphometric and genetic analysis differentiate Mesoamerican population of the endangered stingless bee Melipona beecheii (Hymenoptera: Meliponidae) and support their conservation as two separate units. J. Insect Conserv. 16: 723-731.
Mendes, M.F.M., T.M. Francoy, P. Nunes-Silva, C. Menezes and V.L. Imperatriz-Fonseca. 2007. Intra-populational variability in Nannotrigona testaceicornis Lepeletier, 1836 (Hymenoptera, Meliponini) using relative warps analysis. Bioscience 23: 147-152.
Michener, C.D. 2007. The Bees of the World. Johns Hopkins Press, Baltimore, MD, USA. 913 pp .
Michener, C.D. and S. Boongird. 2004. A new species of Trigona from peninsular Thailand (Hymenoptera: Apidae: Meliponini). J. Kans. Entomol. Soc. 77: 143-146.
Moraes, E.M., V.L. Spressola, P.R.R. Prado, L.F. Costa and F.M. Sene. 2004. Divergence in wing morphology among sibling species of the Drosophila buzzatii cluster. J. Zool. Syst. Evol. Res. 42: 154-158.
Moure, J.M., D. Urban and G.A.R. Melo. 2007. Catalogue of Bees (Hymenoptera, Apoidea) in the Neotropical Region. Sociedade Brasileira de Entomologia. Curitiba. 1058 p. Özkan, A., M.M. Gharleko, B. Özden and I. Kandemir. 2009. Multivariate morphometric study on Apis florea distribution in Iran. Turk. J. Zool. 33: 93-102.

Pavlinov, I.Y. 2001. Geometric morphometrics, a new analytical approach to comparison of digitized images. Zool. J. Moscow. 79: 1-27.
Potts, S.G., J.C. Biesmeijer, C. Kremen, P. Neumann, O. Schweiger and W.E. Kunin. 2010. Global pollinator declines: trends, impacts and drivers. Trends Ecol. Evol. 25:

345-353.
Quezada-Euán, J.J.G., W.J. May-Itzá, M. Rincón, P.D.L. Rúa and R.J. Paxton. 2012. Genetic and phenotypic differentiation in endemic Scaptotrigona hellwegeri (Apidae: Meliponini): implication for the conservation of stingless bees population in contrasting environments. Insect Conserv. Diver. 5: 433-443.
Radloff, S.E., H.R. Hepburn, S. Fuchs, G.W. Otis, S. Hadisoesilo, C. Hepburn and T. Ken. 2005. Multivariate morphometric analysis of the Apis cerana populations of oceanic Asia. Apidologie 36: 475-492.
Ramirez, S.R., J.C. Nieh, T.B. Quental, D.W. Roubik, V.L. Imperatriz-Fonseca and N.E. Pierce. 2010. A molecular phylogeny of the stingless bee genus Melipona (Hymenoptera: Apidae). Mol. Phylo. Evol. 56: 519-525.
Rasmussen, C. and S.A. Cameron. 2010. Global stingless bee phylogeny supports ancient divergence, vicariance, and long distance dispersal. Biol. J. Linnean Soc. 99: 206232.

Rattanawannee, A., C. Chanchao and S. Wongsiri. 2007. Morphometric and genetic variation of small dwarf honeybees Apis andreniformis Smith, 1858 in Thailand. Insect Sci. 14: 451-460.
Rattanawannee, A., C. Chanchao and S. Wongsiri. 2010. Gender and species identification of four native honey bees (Apidae: Apis) in Thailand based on wing morphometic analysis. Ann. Entomol. Soc. Am. 103: 965-970.
Rattanawannee, A., C. Chanchao and S. Wongsiri. 2012. Geometric morphometric analysis of giant honeybee (Apis dorsata Fabricius, 1793) populations in Thailand. J. Asia Pac. Entomol. 15: 611-618.
Rolhf, F.J. 2005a. tpsDig, 2.04 ed. Department of Ecology and Evolution, State University of New York. Stony Brook, NY, USA.
Rolhf, F.J. 2005b. tpsRelw, 2.04 ed. Department of Ecology and Evolution, State University of

New York. Stony Brook, NY, USA.
Sakagami, S.F. 1978. Tetragonula stingless bees of Continental Asia and Sri Lanka (Hymenoptera, Apidae). Fac. Sci. Hokkaido Univ. Ser. VI Zool. 21: 165-247.
Sakagami, S.F., T. Inoue and S. Salmah. 1985. Key to the stingless bee species found or expected from Sumatra, pp. 37-43. In R.I. Ohgushi, (ed.). Evolution Ecology of Insect in Humid Tropics, Especially in Central Sumatra. Kanasawa University Press. Ishikawa, Japan.
Sakagami, S.F. and S.G. Khoo. 1987. Taxonomic status of the Malaysian stingless bee Trigona reepeni, with discovery of Trigona pagdeni from northern Malaya. Kontyû 55: 207214.

Schwarz, H.F. 1939. The Indo-Malayan species of Trigona. Bull. Am. Mus. Nat. Hist. 76: 83-141.
Slaa, E.J., L.A.S. Chaves, K.S. Malagodi-Braga and F.E. Hotstede. 2006. Stingless bees in applied pollination: Practice and perspective. Apidologie 37: 293-315.
Slaa, E.J., L.A. Sanchez, M. Sandi and W. Salazar. 2000. A Scientific note on the use of stingless bees for commercial pollination in enclosures. Apidologie 37: 293-315.
Starr, C.K. and S.F. Sakagami. 1987. An extraordinary concentration of stingless bee colonies in the Philippines with notes on nest structure (Hymenoptera: Apidae: Trigona spp.). Insect Soc. 34: 96-107.
Theeraapisakkun, M., S. Klinbunga and S. Sittipraneed. 2010. Development of a speciesdiagnotic marker and its application for population genetics studies of the stingless bee Trigona collina in Thailand. Genet. Mol. Res. 9: 919-930.
Thummajitsakul, S., S. Klinbunga and S. Sittipraneed. 2010. Development of a speciesdiagnostic marker for identification of the stingless bee Trigona pagdeni in Thailand. Biochem. Genet. 48: 181-192.

Tofilski, A. 2008. Using geometric morphometrics and standard morphometry to discriminate three honeybee subspecies. Apidologie 39: 558-563.
Villemant, C., G. Simbolotti and M. Kenis 2007. Discrimination of Eubazus (Hymenoptera, Braconidae) sibling species using geometric morphometrics analysis of wing venation. Syst. Entomol. 32: 625-634.
Whitfield, C.W., S.K. Behura, S.H. Berlocher, A.G. Clark, J.S. Johnston, W.S. Sheppard, D.R. Smith, A.V. Suarez, D. Weaver and N.D. Tsutsui. 2006. Thrice out of Africa: Ancient and recent expansion of honey bee, Apis mellifera. Science 314: 642-645.

Williams, N.M., E.E. Crone, T.H. Roulston, R.L. Minckley, L. Packer and S.G. Potts. 2010. Ecological and life-history traits predict bee species responses to environmental disturbance. Biol. Conserv. 143: 22802291.

Winfree, R., R. Aguilar, D.P. Vásquez, G. LeBuhn and M.A. Aizen. 2009. A meta-analysis of bees' responses to anthropogenic disturbances. Ecology 90: 2068-2076.


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